

NTU Journal of Agricultural and Veterinary Science - (NTU-JAVS)

NU

(P-ISSN: 2788-9890 e-ISSN: 2788-9904)

Infection Rate of Sarcocystis Spp. of the Intermediate Hosts in Iraq from 1999-2020

1st Huda Ghanim Dakhil 1

Email: 1. <u>hudaalsabahi83@gmail.com</u>
1. Ministry of Education -Gifted school .

Received: 13-4-2022, Accepted: 16-10-2022, Published online: 10-11-2022

Abstract. Sarcocystis spp. are protozoal parasites that are widely spread in mammals, reptiles, and birds. The disease they cause is called Sarcocystosis. The aim of this review is to gain knowledge of the infection rate and species of Sarcocystis in intermediate hosts in the provinces of Iraq and the areas that haven't been notified till now. Thirty relevant published research were collected about Sarcocystis spp. that infected the intermediate hosts in Iraq. Ten species determined in some intermediate hosts in Iraq are only: S. tenella, S. arieticanis, S. gigantean, S. ovicanis, S. capricanis, S. caprifelis, S. moulei, S. cruzi, S. cameli and S. fusiformis. There have been no pupulation studies on Sarcocystis in the following Iraqi provinces: Kurkuk, Saladin, Diyala, Wasit, Al-Qadisiya, Al-Najef Al-Ashraf, Dhi Qar and Al-Muthanna. The infection rate ranged from 0.1% in Missan in water buffalo to 100% in several animals in Nineveh, Baghdad and Duhok of Iraqi provinces. The Sarcocystis spp. can be diagnosed by several methods.

Keywords: *Sarcocystis,* intermediate hosts, Iraq. **Note**: The present study is a review literature study.

Introduction

Sarcocystis spp. are worldwide distributed parasites in human beings and several species of animals. They infect different muscles of the body. The Sarcocystis protozoan parasite (Phylum: Apicomplexa, Family Sarcocystidae) (Fayer, 2004). A sexual stage and cyst formation take place in the intermediate host (human beings and several species of animals) . Sexual stageand formation oocysts take place in the definitive host (human beings and predators such as dogs or cats). The definitive host gets infected when it eats meat containsing encysted parasites. Each encysted contains hundreds to thousands of Sarcocystis bradyzoites. In the intestinal wall of the definitive host, each bradyzoite directly develops into a microgamete or macrogamete, which fertilizes to form oocysts. Mature oocysts which have two sporocysts are eleminated with the feces. Intermediate hosts will be infected when they eat vegetables or drink water contaminated with oocysts (or sporocysts). The ingested oocysts or sporocysts cross into the blood stream. They can multiply in the small blood vessels before invading the different muscles where they eventually develop into bradyzoites, and mature into

sarcocyst usually taking about 2 months or more (Saeed *et al.*, 2018) (See Fig. 1).

Sarcocystis species may cause several symptoms such as diarrhea, vomition and abdominal pain (Koneman et al., 2005; Juckett, 1996). Death occurs in sever cases due to electrolytes and water loss (Koneman et al., 2005). The muscular infections are possible causes of some cardiac diseases (viz. myocarditis, cardiomyopathy or valvular lesions) and rheumatic (viz. imyositis and musculoskeletal complaints) (Habeeb et al., 1996). Sarcocystis spp. infection in the intermediate mammals may cause abortion, but we don't know if this can happen in human cases (Greve, 1985). Treatment of intestinal sarcocystosis are usually

self-limiting (Bunyartvej *et al.*, 1982). Prevention of infections in intermediate hosts depends on reducing exposure to oocysts or sporocysts. And in the definitive host, it is based on preventing eating of raw or not well cooked animal tissues. Freezing raw meat, to -20°C for about 48 hours or -4°C for about 48 hours, can be effective on parasites and can prevent infection because it will destroy the parasites (Leek *et al.*, 1978; Aiello and Moses, 2016).

The aims of the present study are to record the infection rate of *Sarcocystis* spp. in intermediate

hosts in different regions of Iraq and the areas that have not been registered till now, as well as, to urge other researchers to study this disease in all Iraqi areas to form an imagination about this epidemic and the species of *Sarcocystis* in Iraq. In

addition, the study aims to increase studies on sarcocystosis specially in Kurkuk, Saladin, Diyala, Wasit, Al-Qadisiya, Al-Najef Al-Ashraf, Dhi-Qar and Al-Muthanna to diagnose many other types of *Sarcocystis*.

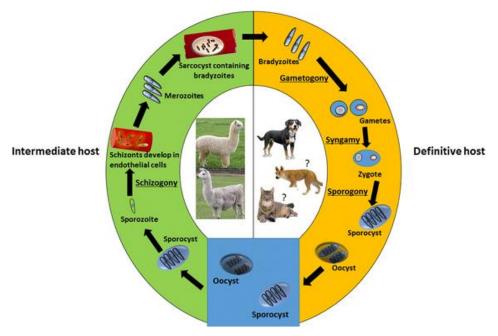


Fig 1. Life cycle of Sarcocystis spp (Saeed et al., 2018).

Method

A total of thirty relevant research (English and Arabic) from PubMed, Google Scholar and articles dealing with *Sarcocystis* spp. published in Iraqi Academic Scientific Journals from 1999-2020 were used to provide the present study the rates of infections of this disease in several intermidiate hosts in 10 Iraqi provinces including: Nineveh, Sulaimania, Duhuk, Erbil, Baghdad, Karbala, Babylon, Anbar, Missan and Basra. Data from these references were collected to show *the Sarcocystis* spp. study areas, hosts, species of

Sarcocystis, methods of diagnosis and infection rates. The arrangment of the Table (1) depends on the publication years of these research starting from the oldest down to the newest.

Literature Review

As mentioned before, thirty relevant research on *Sarcocystis spp.* was collected and shown in Table (1), as well as, all the species infectioneding the intermidiate hosts in several Iraqi provinces shown in Table (2) to facilitate the review studies.

Provinces	Infections	Species of	Examination	Infection rate	References
	animals	Sarcosystis	Methods	%	
'	Sheep		Pepttic digestion method	93.3	
	Goats	Sarcosystis spp.	Indirect fluorescent antibody	88.6	
Baghdad	Cattle	(Macroscopic	test (IFAT)		(Latif et al., 1999)
	Water buffloes	and microscopic)	Squeezing	81.3	
	Camels		Muscle squash	81.2	
			1-Macroscopical	33.6	
		S. caprifelis	examination		
Sulaimania	Goats	S. capracanis	2-Microscopical examination	97.4	(Barham et al., 2005)
			(Muscle squash, Pepsin		
Northern Iraq	Goats	S.capracanis	digest of muscles, Histology) Macroscopic examination	34	(Barham et al., 2005)
Northerniaq	Guais	o.capracariis	Macroscopic examination	J 4	(Bamam et al., 2003)
Nineveh	Sheep	S. gigantea	Histopathological	94.83	(Al-Taee and Al-Hyali , 2007) (in Arabic)

			1-gross exam 2-trichinoscopy		
Dohuk	Goats	Sarcosystis spp.	3-squeezing method	93%	(Al-Bayati, 2008)
			4- post trichinoscopy		
			5-drop exam 1-gross examination		
		Sarcosystis spp.	2-trichinoscopy		(Al-Nakshabandi,
Dohuk	Cattle	(microscopic	3-squeezing method	81-96	2008)
		cysts)	4-post trichinoscopy drop examination		_000)
	Sheep		ELISA	0.0	
	Goats		IFAT	82 60	(Al-Taee et al., 2009)
Nineveh	Cattle	Sarcosystis spp.	IHAT	45	(in Arabic)
	Human		Agar Gel Immunodiffusion Test	26	,
			1-Trichinoscopy	63.63	
Dohuk	Sheep	Sarcosystis spp.	2-Post trichinoscopy drop	81.81	(Barwary et al.,
			3-Peptic digestion 4-histopathlogical	100 100	2009)
			Pepttic digestion method	100	/Makanana alamat
Babylon	Goats	S. caprifelis	Trichenoscopy	0.39	(Mohammed and Kadihm, 2010) (in
Dabyion	Coalo	S. capracanis	Squeezing	93.22	Arabic)
		Microscopic	Histopathological Histopathological		(Al-Sadi& younis,
Nineveh	Sheep	Sarcosystis spp	i notopati iologica.	100	2010)
Nineveh	Sheep	S. gigantea	Macroscopic examination		(Hyali et al., 2011)
	•		Histological test 1-Macroscopic examination		
Anbar	Camels	S. cameli	2-muscle ingestion with	1.78	(Shahatha, 2011) (in
		•	pepsin and fluid testing	80.3	Arabic)
Duhok	Sheep Goats	S. gigantea S. caprifelis	Macroscopic examination	7 8	(Al-Bayati,2012)
5.1.1	Slaughtered	•	Trichenoscopy	66.5	(1.4.1
Babylon	Cattle	S.cruzi	histological techniques	70	(Mohammad, 2012)
			Macroscopic examination		
Babylon	Sheep	S. gigantea	muscle ingestion with pepsin and fluid testing	7.69	(Mohammed and
2007.0	ооор	S. tenella	Trichenoscopy	97.93	Kadihm, 2013)
			Histological test		(11(-1, 0044)
Nineveh	Pigeons	Sarcocystis spp.	Trichenoscopy	3.08%	(Hasan <i>et al</i> ., 2014) (in Arabic)
Nineveh	Choon	Caracayatia ann	Histologicaltechniques to	1%	(kako and Alhbiti,
Mineven	Sheep	Sarcocystis spp.	brain		2014)
Basra	Buffalo		Trichenoscopy Squeezing	77.5 72.2	(Maktoofa and abeekh, 2015)
Dawkdad	Chaan	S.tenella	Digestion method		, ,
Baghdad	Sheep		PČR	100	Whaeeb <i>et al.</i> , 2016)
Nineveh	Sheep	S. gigantea	Macroscopic examination	94.8	(Al-Taee& Al-Hayali, 2017) (in Arabic)
			1-Direct Scotch Cellophane		2017) (1117(10010)
			Adhesive Tape		
Duhok	Sheep	S.ovicanis	2-Muscle Mincing and Squash Method	97.5	(Zangana and
Dullok	Goats	S.capricanis	3-Acid pepsin Digestion Test	100	Hussein, 2017)
			4-Histopathological		
		0 6 16 1-	Examination	0.0	
Missan	Water buffloes	S.fusiformis S. moulei	Macroscopic examination PCR	2.8 0.1	(Dakhil et al., 2017)
	Human :	JJuloi	. •	33.33	
	1-Rheumatoid			33.33	
Missan	arthritis patients 2-non-	Sarcocystis spp.	ELISA		(Dakhil et al., 2017)
	heumatoid			00.0	
	arthritis patients	_		23.8	
Nineveh	Pigeon	Sarcocystis spp. S.tenella and S.	Trichinoscopy Transmission Electron	3.08	(Hasan et al., 2018)
Baghdad	Sheep	arieticanis	Microscopic		(Whaeeb, 2018)
-			<u>ı</u>		

Nineveh	Sheep	Sarcocystis spp.	Histopathological	35.5	(Hamdany, 2019)
Erbil	Sheep Goats	S. gigantea S. moulei	Macroscopic examination Histopathological Examination	9.5 8.8	Swar and Shnawa, 2020)
Baghdad	Slaughtered Cattle	S. cruzi	PCR	2.5 (depended on primar)	(Kamil and Faraj, 2020)
Karbala	Sheep	Sarcocystis spp.	Macroscopic examination Microscopic examination: 1-Acid pepsin Digestion 2-Meat mincing by garlic presse 3-PCR	5.82 90.78	(Al-Saadi <i>et al.</i> , 2020)
Duhok	Cattle	Sarcocystis spp.	Macroscopic examination Microscopic examination: 1-tissue squeezing 2-muscle squash 3-pepsin hydrochloric aacid digestion 4-histopathological examination	76	(Mohammad <i>et al.</i> , 2020)

Table 1. The record Sarcocystis spp. in Iraqi provinces.

Infection intermidate hosts	Sarcocystis spp.
Human	Sarcosystis spp.
	S.tenella
	S. arieticanis
	S. gigantean
Sheep	S.ovicanis
Sileep	S.capricanis
	S. caprifelis
	S. moulei
	S. caprifelis
	S. capracanis
Conto	S.ovicanis
Goats	S. moulei
	S. gigantean
Cattle	S. cruzi
Camels	S. cameli
Matarbufflaga	S.fusiformis
Water buffloes	S. moulei
Pigoen	Sarcocystis spp.

Discussion

The present study is the first literature review stydy dealing with the infection rate of Sarcocystis spp. of the intermediate hosts in Iraq. Latif et al (1999) is the oldest research collected in this review. It recorded that Sarcocystis spp. in goats, sheep, water buffaloes, cattle and camels in Iraq use different methods to diagnose it. Besides, the study observed several methods for diagnosing Sarcocystis in Iragi research including: macroscopical examination to see the cysts by (Macrocystes in muscles), microscopical examination methods which are Pepttic digestion method, indirect fluorescent antibody test (IFAT), squeezing muscle squash, histology, trichinoscopy

and PCR (Barham *et al.*, 2005; Al-Saadi *et al.*, 2020). These methods were the same methods used in several research in the world. Stojecki *et al.* (2012) used PCR, histological examination (Meistro *et al.*, 2015), and transmission electron microscope (Jehle, 2009).

Ten species were only determined in some intermediate hosts in Iraq which were as follows: S.tenella, S. arieticanis, S. gigantean, S.ovicanis, S.capricanis, S. caprifelis, S. moulei, S. cruzi, S. cameli and S.fusiformis. On the other hand, there were several species diagnosed in the world in addition to the species recorded in this review study: S. hirsuta, S. hominis in bovine (Meistro et al., 2015), S. tenella (Dong et al., 2018), S.

medusiformis in sheep (Obendorf and Munday 1987), S.bertrami and S. fayeri in horse (Chun-Li Ma et al., 2020).

There are no studies on *Sarcocystis* in the following Iraqi provinces; viz. Kurkuk, Saladin, Diyala, Wasit, Al-Qadisiya, Al-Najef Al-Ashraf, Dhi-Qar and Al-Muthanna. There were no clear reasons preventing the researchers from studying this parasite in these provinces. The present study noticed the infection rate ranged from 0.1% in Missan to 100% in Bagdad, Nineveh, Dohuk provinces (See Table 1).

This different rate can be due to the differences that existed in definative hosts, such as dogs, cats, etc. Or, it may be related to the weakness of sanitary control in the slaughter houses, as it requires to distruct infected meat. Eating undercooked infected meat is one of the most important factors for human beinng's infection. Thus, human can become a definative host when eating undercooked meat (Beaver *et al.*, 1979).

Conclusion

Finally, the study concludes the following:

- 1-Several infections were recorded in many intermediate hosts in different Iraqi areas.
- 2- Sarcocystis spp. can be diagnosed by several methods including: peptic digestion, indirect fluorescent antibody test (IFAT), squeezing muscle squash, histology, trichinoscopy, PCR. and transmission electron microscope.
- 3- In 44% of Iraqi places and cities, there were no manuscript worked to daignosing *Sarcocytis*.

References

- [1] Aiello, S. E. and Moses, M. A., (2016) The Merck veterinary manual. 11th ed. Kenilworth, NJ: Merck and Co; Equine protozoal myeloencephalitis. 10-1309.
- [2] Al- Bayati, S. M. H. (2012) Studying some biochemical parameters of *Sarcocystis* parasites isolated from local sheep and goats in Duhok area. Al-Anbar J. Vet. Sci. 5 (2): 94-97.
- [3] Al-Bayati, S. M. (2008). Comparative study on goats sarcocystosis diagnostic techniques in Dohuk governorat. Journal of university of Anbar for Pure science. 2 (2): 1-5.
- [4] Al-Hamdany, E. Kh.(2019) Pathological study of some esophageal lesions of slaughtered sheep in Mosul abattoir. *raqi Journal of Veterinary Sciences*. 34(1): 145-151.
- [5] Al-Hasnawy, M. H. (2012) Prevalence of bovine sarcocystosis in Babylon province. Kufa Hournal For Veterinary Medical Sciences. 3(2):78-83.
- [6] Al-Hyali, N. S. , Kennany, E. R. and Khalil, L.Y.(2011). Fate of macrosarcocyst of *Sarcocystis gigantea*in sheep. Iraqi Journal of Veterinary Sciences. 25(2): 87-91.

- [7] Al-Nakshabandi, A. (2008). Rative study on cattle sarcsystosis diagnostic techniques in dohuk governorate. Basrah Journal of Veterinary Research. 7(2): 16-20.
- [8] Al-Nakshabandi, A. (2008) Rative study on cattle Sarcocystosis diagnostic techniques in Duhok governorate. Bas J Vet Res. 7(2):16-20.
- [9] Al-Saadi, S. A. M., Al-Mussawi, K. A. M. and Muhammed, H. A (2020) Molecular Identification of *Sarcocystis* Species Infection in Sheep in *Karbala* Governorate-Iraq. Medico-legal Update, January-March. 20 (1): 889-895.
- [10] Al-Sadi, H. I. and Younis, H. B. (2010) Orallesion in sheep in Mosul area, Iraq. Veterinary Research . 3(2): 21-26.
- [11] Al-Taee, A. F. and Al-Hyali, N. S. (2007) Toxicity of Sarcocytis gigantea detected From sheep in ninevah governorate. Iraqi Journal of Veterinary Sciences. 21(2): 229-243.
- [12] Al-Taee, A. F., Al-Hyali, N. S. and Al-Badree, M. S. (2009) Seroprevalence of antibodies against Sarcocystis gigantea in different hosts in Ninevah governorate. Iraqi J. Vet. Sci.2009; 23 (1): 107– 112.
- [13] Bahram, M. M., Syutzer, H., Karanis, P., Latif, M. P. and Neiss, W. F. (2005) Seasonal variation in *Sarcocystis* species infections in goats in northern Iraq. Parasitology ·130: 151-6.
- [14] Barwary, S. Q and Al-Bayati, S. M.(2009) Trocarization for diagnosis of sarcocystosis in sheep Bas.J.Vet.Res. 8(1):78-87.
- [15] Bunyarativej, S. and Bunyawongwongwiroj, P., Nitiyanant, (1982) Human intestinal sarcosporidiosis: report of six cases. AM. J. Trop. Med. Hyg. 31: 36-41.
- [16] Chun –Li , Yu-Long, Tao Wen, et al. (2020) Prevalence and morphological and molecular characteristics of *Sarcocystis bertrami* in horses in China . Parasite 27(1):1-7.
- [17] Dakhil, H. G., Abdallah, B. H. and Abdallah, F. A. (2017a) Molecular identification of *Sarcocystis fusiformis* and *S. moulei* infecting water buffaloes (*Bubalus bubalis*) in southern Iraq. World Journal of Pharmaceutical Research. 6: 215-229.
- [18] Dakhil, H. G., Abdallah, B. H. And Abdallah, F. A. (2017B) *Sarcocystis* spp. in relation to non-specific and rheumatoid arthritis diseases. World Journal of Pharmaceutical Research. 6 (2): 1298-1308.
- [19] Dong, Hui , Ruijing, Su, et al (2018) Sarcocystis species in wild and domestic sheep (Ovis ammon and Ovis aries) from China. BMC Veterinary Research 14:377.
- [20] Faraj, A. A., Kawan, M. H. (2012) Detection of Sarcocystosis in some wild birds. Iraqi J.Vet. Med. 36: 56-70.
- [21] Fayer, R., Douglas, H. E. and Dubey, J.(2015). Human infection with *Sarcocystis* species. American Society for Microbiology. 28(2): 295-311.
- [22] Fayer, R. (2004) Sarcocystis spp. In human infections. Clinical Microbiology Reviews, 17(4): 894-902.

- [23] Greve, E. (1985) Sarcosporidiosis, overlooked zoonosis. Man as intermediate and final host. Dan Med Bull. 32(4): 228-30.
- [24] Habeeb, Y. S., Selim, M. A., Ali, M.S., Mahmoud L. A., Abdel Hadi AM and Shafei A. .(1996) Serological diagnosis of extraintestinal Sarcocystosis. J Egypt Soc Parasitol. 26 (2): 393-400.
- [25] Hasan, M. H. (2018) Al AbbadiA E, Abdul Ruhman N R. A Study of Endoparasites of Pigeons in Mosul City. Rafidain journal of science. 27(1A): 76-81.
- [26] Juckett , G.(1996) Intestinal protozoa. Am Fam Physician. 53(8):2507-18.
- [27] kako, M. D. And Alhbiti T. H.Y. (2014) Prevalence of brain lesions in sheep slauchtered Mosul area during 2012-2013. Basrah Journal of Veterinary Research. 13(1): 284-302.
- [28] Kamil, J.K. and Faraj, A. A. (2020) Identification of Sarcocystis spp. in imported deef by traditional and molecular technique .. Plant Archives Vol. 20, Supplement 2, pp. 25-36.
- [29] Koneman, E. W., Allen, S. D. and Janda, W. M (2005) Schrechenberg PC and Winn WC. Color atlas and textbook of diagnostic microbiology. Philadelphia: J B Lippincot Company (sisth edition). 1736p.
- [30] Latif, B. M., Al-Delemi, J. K., Mohammed, B. S., Al-Bayati, S. M. and Al-Amiry, A. M.(1999). Prevalence of *Sarcocystis* spp. in meat-producing animals in Iraq. Vet Parasitol. 84(1-2):85-90.
- [31] Leek, R. and Fayer, R. (1978). Infectivity of *Sarcocystis* in beef and beef products from a retail food store. Proc. Helminthol. Sos.Wash.45:135-136
- [32] Maktoofa, A. R. and Tabeekh, M. A. S. (2015). *Sarcocystis* infection buffalo carcassesin Basra govervorate. Iraq. International Journal of Development Research. 5(11): 6025-6028.
- [33] Meistro, S., Peletto, S., Pezzolato, M., Varello, K., Botta, M. and Richelmi, G. (2015) *Sarcocystis* spp. prevalence in bovine minced meat: a histological and molecular study. Ilalian Journal of Food Safety. 4(2): 85-87.
- [34] Mohamed, A. and Hussein S. N., Shukur, M. S., Mohammad, R. A., Ali, A. A. and Khalil, L, N. (2020) Survey on *Sarcocystis* infection in imported male cattle carcasses slaughtered at Duhok abattoir, Kurdistan region of Iraq Teroj. Microbial Biosystems. 5(1): 128-134.
- [35] Mohammad, M. H. and kadihm, F. S.(2010) Epidemiological Study of caprine *Sarcocystosis* in Babylon province Al-Qadisiyah Journal of Veterinary Medicine Sciences. 9(2): 5-15.
- [36] Mohammed, H. M. (2013) Study in prevelance of sarcocystis in sheep in Babylon.
- [37] Muhamed, T. A. and Al-Barwary, L. T. O. (2016) Prevalence of Intestinal Parasites in the Intestine of Dogs (SheepKeeper, Owned, Pet and Stray) in Duhok Province. Kurdistan Region, JVST. 7(6):379.
- [38] Obendorf, D. L. and Munday, B. L. (1987) Experimental infection with *Sarcocystis medusiformis* in sheep. Veterinary Parasitology, 24(1-2): 59-65.

- [39] Saeed, M. A. and Rashid, M. H. (2018) Vaughan J, Abdul Jabbar. Sarcocystosis in south American camelids: The state of play revisited. Parasites&Vectors. 11:146.
- [40] Shahatha, S. S. and Al-Karboly, R. W. K. (2011) The sarcosystosis caused by *Sarcocystis cameli* infestation prevalence among camels in Al-Anbar governorate –Iraq. Iraqi Journal of Desert studies. 3(1): 186-191.
- [41] Spickler Anna Rovid. (2020) Sarcocystosis. Retrieved from http://www.cfsph.iastate.edu/DiseaseInfo/factshe ets.php.)) edd to reference in research im treatment.
- [42] Stojecki, K., Karamon, J., Sroka, j. and Cencek, T.(2012) Molecular dignostics of *Sarcocystis* spp. infection. Polish Journal of Veterinary Sciences . 15(3)589-596.
- [43] Swarand, S. O. and Shnawa, B. H. (2020) Ultra structural and Molecular Characterization of Sarcocystis species Derived from Macroscopic Sarcocystis of Domestic Sheep and Goats in Soran City, Erbil, Iraq. World Vet J. 10 (4): 540-550.
- [44] Whaeeb, S. T. and Faraj, A. A.(2018) Identification of *Sarcocystis* spp. by Transmission Electron Microscope. International Journal of Science and Research (IJSR). 7(2): 357–359.
- [45] Whaeeb, S. T. and Faraj, A. A. (2016) Molecular identification and phylogeny of microscopic *Sarcocystis* sheep in Baghdad Province. International Journal of Advanced Research in Biological Sciences. 3(12): 50-56.
- [46] Zangana, I. K. and Hussein, S. N. (2017) Prevalence of Sarcocystis Species (Sarcocystis ovicanis and Sarcocystis capricanis) in Tongue Muscle of Sheep and Goats in Duhok Province, Kurdistan Region, North Iraq. The Scientific Journal of Koya University. 5(1): 36-40. 36-4.